



Ontologies & CCF: Anatomical Structures, Cell Types, and Biomarkers

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*“External Collaborations” Session @ HuBMAP All-Hands Meeting
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What is a CCF?

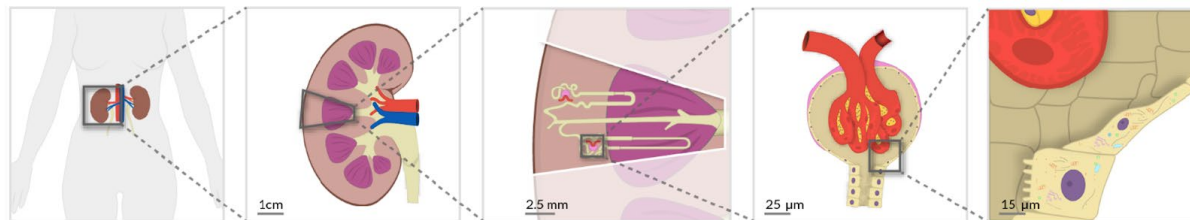
The Common Coordinate System (CCF) consists of ontologies and reference object libraries, computer software, e.g., user interfaces, and training materials that

- enable biomedical experts to semantically annotate tissue samples and to precisely describe their locations in the human body (“registration”),
- align multi-modal tissue data extracted from different individuals to a reference coordinate system (“mapping”) and,
- provide tools for searching and browsing HuBMAP data at multiple levels, from the whole body down to single cells (“exploration”).

CCF Requirements

The CCF must capture major **anatomical structures, cell types, and biomarkers** and their interrelations across **multiple levels of resolution**.

It should be **semantically explicit** (using existing ontologies, e.g., Uberon, CL) and **spatially explicit** (e.g., using 3D reference organs for registration and exploration).



Body

- Body
- Kidney (Left, Right)
- Aorta
- Renal artery
- Renal vein
- Ureter

Organ

- Renal capsule
- Renal pyramid
- Renal cortex
- Renal medulla
- Renal calyx
- Renal pelvis

Functional Tissue Unit

- Nephron
- Renal corpuscle
- Proximal convoluted tubule
- Loop of Henle
- Distal convoluted tubule
- Connecting tubule
- Collecting duct

FTU Sub-structure(s)

- Bowman's capsule
- Glomerulus
- Efferent arteriole
- Afferent arteriole

Cellular

- Parietal epithelial cell
- Capillary endothelial cell
- Mesangial cell
- Podocyte

ASCT Tables

Anatomical Structures and Cell Types (ASCT) tables aim to capture the partonomy of anatomical structures, cell types, and major biomarkers (genomic, epigenomic, transcriptomic, proteomic, lipidomic, and metabolomic).

Structure/Region	Substructure/Sub region	Cell Type	Subset of Marker Genes
Renal Corpuscle	Bowman's Capsule	Parietal epithelial cell	<i>CRB2*</i> , <i>CLDN1*</i>
	Glomerulus	Podocyte	<i>NPHS2*</i> , <i>PODXL*</i> , <i>NPHS1*</i>
		Capillary Endothelial Cell	<i>EHD3*</i> , <i>EMCN*</i> , <i>HECW2*</i> , <i>FLT1*</i> , <i>AQP1*</i>
		Mesangial Cell	<i>POSTN*</i> , <i>PIEZO2*</i> , <i>ROBO1*</i> , <i>ITGA8*</i>

Partial ASCT Table from

- El-Achkar et al. A Multimodal and Integrated Approach to Interrogate Human Kidney Biopsies with Rigor and Reproducibility: The Kidney Precision Medicine Project. bioRxiv. 2019; 828665. doi:10.1101/828665

Table 3: Cell types and associated markers from KPMP Pilot 1 transcriptomic studies. Asterisk denotes genes detected by more than one technology. *Italics*, genes detected by a single technology.

Structure/R region	Sub structure/Sub region	Cell Type	Abbreviation	Subset of Marker Genes	Pertinent negatives/comments
Renal Corpuscle	Bowman's Capsule	Parietal epithelial cell	PEC	<i>CRB2*</i> , <i>CLDN1*</i>	
	Glomerulus	Podocyte	POD	<i>NPHS2*</i> , <i>PODXL*</i> , <i>NPHS1*</i>	
		Capillary Endothelial Cell	GC-EC	<i>EHD3*</i> , <i>EMCN*</i> , <i>HECW2*</i> , <i>FLT1*</i> , <i>AQP1*</i>	
		Mesangial Cell	MC	<i>POSTN*</i> , <i>PIEZO2*</i> , <i>ROBO1*</i> , <i>ITGA8*</i>	
Tubules	Proximal Tubule	Proximal Tubule Epithelial Cell (general)	PT	<i>CUBN*</i> , <i>LRP2*</i> , <i>SLC13A1*</i> , <i>ALDOB*</i> , <i>GATM*</i>	There is overlap among the segments
		Proximal Convoluted Tubule Epithelial Cell Segment 1	PT-S1	<i>SLC5A2*</i> , <i>SLC5A12*</i>	
		Proximal Tubule Epithelial Cell Segment 2	PT-S2	<i>SLC22A6*</i>	
		Proximal Tubule Cell Epithelial Segment 3	PT-S3	<i>PDZK1IP1*</i> , <i>MT1G*</i>	
	Loop of Henle, Thin Limb	Descending Thin Limb Cell (general)	DTL	<i>CRYAB*</i> , <i>VCAM1*</i> , <i>AQP1*</i> , <i>SPP1*</i>	<i>CLDN10</i> low
		Ascending Thin Limb Cell (general)	ATL	<i>CRYAB*</i> , <i>TACSTD2*</i> , <i>CLDN3*</i>	<i>AQP1</i> low to none
	Loop of Henle, Thick Limb	Thick Ascending Limb Cell (general)	TAL	<i>SLC12A1*</i> , <i>UMOD*</i>	<i>SLC12A3</i> low to none
		Cortex-TAL cell	C-TAL	<i>SLC12A1*</i> , <i>UMOD*</i>	
		Medulla-TAL cell	M-TAL	<i>SLC12A1*</i> , <i>UMOD*</i>	
		TAL-Macula <i>Deposa</i> cell	TAL_MD	<i>NOS1*</i> , <i>SLC12A1*</i>	
Distal Convolution	Distal Convoluted Tubule Cell (general)	DCT	<i>SLC12A3*</i> , <i>TRPM6*</i>		
	DCT type 1 cell	DCT-1	<i>SLC12A3*</i> , <i>TRPM6</i>	<i>SLC8A1</i> , <i>HSD11B2</i> (low to none)	
	DCT type 2 cell	DCT-2	<i>SLC12A3*</i> , <i>SLC8A1*</i> , <i>HSD11B2</i>	Has CNT and DCT signature	
	Connecting Tubule	Connecting Tubule Cell (general)	CNT	<i>SLC8A1*</i> , <i>CALB1</i> , <i>TRPV5</i>	<i>SLC12A3</i> low to none. IC or PC without <i>SLC8A1</i> could be in the CNT structure
		CNT-Principal Cell	CNT-PC	<i>SLC8A1*</i> , <i>AQP2*</i> , <i>SCNN1G*</i>	
		CNT-Intercalated Cell	CNT-IC	<i>SLC8A1*</i> , <i>CA2</i> , <i>ATP6VOD2*</i>	
CNT-IC-A cell		CNT-IC-A	<i>SLC8A1*</i> , <i>SLC4A1*</i> , <i>SLC26A7*</i>		
	CNT-IC-B cell	CNT-IC-B	<i>SLC8A1*</i> , <i>SLC26A4*</i> , <i>SLC4A9*</i>		
Collecting Duct	Collecting duct (general) cell	CD	<i>GATA3*</i>	<i>GATA3</i> may be in subpopulation of DCT, CNT and <i>vSMC/P</i> . <i>SLC8A1</i> , <i>CALB1</i> , <i>TRPV5</i>	
	CD-PC (general)	CD-PC			
	C-CD-PC	C-CD-PC	<i>AQP2*</i> , <i>AQP3*</i> , <i>FXYP4*</i>		
	M-CD-PC	M-CD-PC	<i>SCNN1G*</i> , <i>GATA3*</i>		
	Outer medulla-CD-PC	OM-CD-PC			
	Inner Medulla-CD cell	IM-CD	<i>AQP2*</i> , <i>SLC14A2</i>		

Vessels	Endothelial Cells (non-glomerular)	Transitional PC-IC cell	IRC-IC	<i>FXYP4*</i> , <i>SLC4A9*/SLC26A7*</i>	(low to none); Low to No
		CD-IC (general) cell	CD-IC	<i>CA2</i> , <i>ATP6VOD2*</i>	<i>CALCA</i> and <i>KIT</i> in C-CD-IC-A. It may not be possible to assign IC or PC to <i>CNT</i> or CD structures without regional information of their source.
		CD-IC-A (general) cell	CD-IC-A	<i>SLC4A1</i> , <i>SLC26A7*</i> , <i>TMEM213*</i>	
		C-CD-IC-A cell	C-CD-IC-A	<i>SLC26A7*</i> , <i>SLC4A1*</i>	
		M-CD-IC-A cell	M-CD-IC-A	<i>SLC26A7*</i> , <i>SLC4A1*</i> , <i>KIT*</i> , <i>CALCA</i>	
		CD-IC-B (general) cell	CD-IC-B		
		C-CD-IC-B cell	C-CD-IC-B		
		M-CD-IC-B cell	M-CD-IC-B	<i>SLC4A9*</i> , <i>SLC26A4*</i>	
		EC-IC-B cell	EC-IC-B		
		EC-Lymphatics	EC-LYM	<i>MMRN1*</i> , <i>PROX1</i>	
Vessels	Endothelial Cells (non-glomerular)	Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	
		EC-Afferent/Effluent Arteriole	EC-AEA	<i>SERPINE2*</i> , <i>TM4SF1*</i>	likely <i>PALMD</i>
		EC-Peritubular capillaries	EC-PTC	<i>PLVAP*</i>	
		EC-Descending Vasa Recta	EC-DVR	<i>TM4SF1*</i> , <i>PALMD</i>	
		EC-Ascending Vasa Recta	EC-AVR	<i>DNASEIL3*</i>	low to none
Structure/R region	Sub structure/Sub region	Cell Type	Abbreviation	Subset of Marker Genes	Pertinent negatives/comments
Interstitial	Stroma (non-glomerular)	Vascular Smooth Muscle/Pericyte (general)	<i>vSMC/P</i>	<i>TAGLN*</i> , <i>ACTA2*</i> , <i>MYH11*</i> , <i>NTRK3</i> , <i>MCAM</i>	
		<i>vSMC/P</i> -Renin	<i>vSMC/P</i> -REN	<i>REN</i>	
Vessels	Immune	Fibroblast	FIB	<i>DCN*</i> , <i>ZEB2</i> , <i>C7</i> , <i>LUM</i>	
		Macrophages-Resident	MAC-R	<i>CD163*</i> , <i>IL7R*</i>	
		Macrophage	MAC	<i>ST00A9</i>	
		Natural Killer Cell	NKC7	<i>NGK7</i>	
		Dendritic Cell	DC	<i>APOE</i>	
		Monocyte	MON	<i>CTQA</i> , <i>HLA-DRA</i>	
		T lymphocyte (general)	T	<i>CD3</i>	
		T Cytotoxic	T-CYT	<i>GZMA</i>	
		B lymphocyte	B	<i>IGJ</i>	

El-Achkar et al. A Multimodal and Integrated Approach to Interrogate Human Kidney Biopsies with Rigor and Reproducibility: The Kidney Precision Medicine Project. bioRxiv. 2019; 828665. doi:10.1101/828665

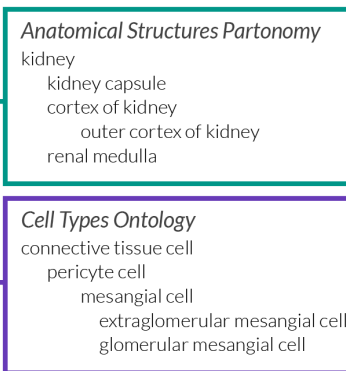
ASCT Table Usage

ASCT tables guide **CCF Ontology** and **3D Reference Object Library** design that semantically name and spatially place tissue data from different individuals into one CCF (i.e., mapping).

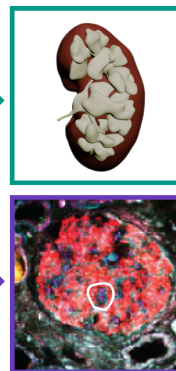
ASCT Table

Structure/Region	Sub structure/Sub region	Cell Type
Renal Corpuscle	Bowman's Capsule	Parietal epithelial Cell
	Glomerulus	Podocyte
Renal Tubule	Proximal Tubule	Mesangial Cell
		Proximal Tubule Epithelial Cell (general)
		Proximal Convoluted Tubule Epithelial Cell Segment 1
		Proximal Tubule Epithelial Cell Segment 2
		Proximal Tubule Epithelial Cell Segment 2
	Loop of Henle, Thin Limb	Descending Thin Limb Cell (general)
		Ascending Thin Limb Cell (general)
	Loop of Henle, Thick Limb	Thick Ascending Limb Cell (general)
		Cortex-TAL Cell
	Distal Convolution	Medulla-TAL Cell
		TAL-Macula Densa Cell
		Distal Convoluted Tubule Cell (general)
Connecting Tubule	DCT Type 1 Cell	
	DCT Type 2 Cell	
	Connecting Tubule Cell (general)	
	CNT-Principal Cell	

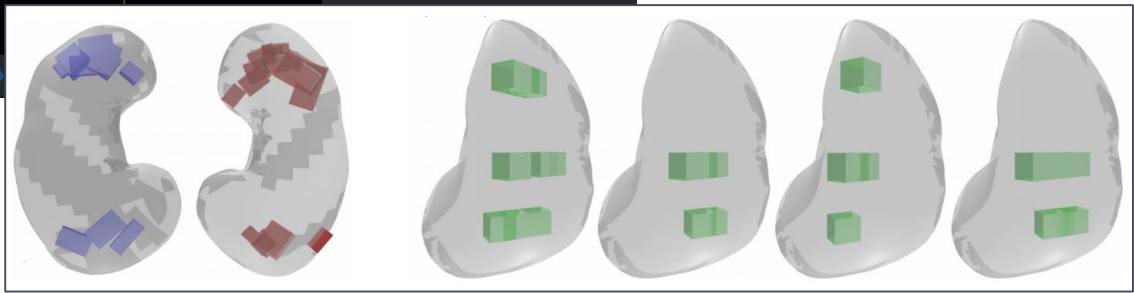
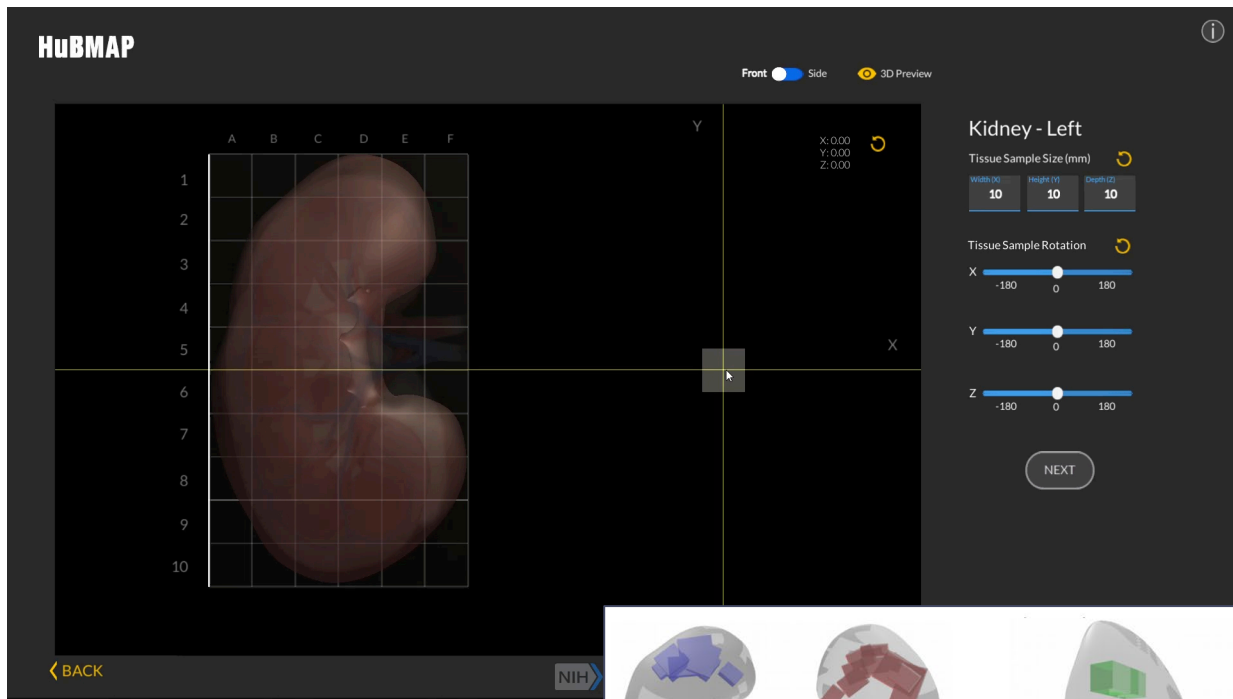
Ontology



3D Reference Object Library




Tissue blocks are registered into the CCF using the Registration User Interface (RUI), and they can be explored via the Exploration User Interface (EUI).



For the first HuBMAP portal release, 48 tissue blocks were registered.

1st Portal Release: Upload Portal



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HuBMAP Display ID Generator

Generate unique identifiers which will be used consortium wide to track sample and associate data with samples.

Source HuBMAP ID * [Look up](#)

HuBMAP display id: **TEST0005-RK**

type: Organ name:

Organ Type: Kidney (Right)

HuBMAP ID: HBM:264-TTJ-798

Description:

Tissue Sample Type *

Protocol 1

protocols.io DOI *

Protocol document * [Browse](#)

doc, docx and pdf files only

[Add Protocol](#)

Generate IDs for multiple FFPE block samples

Lab IDs and Sample Locations can be assigned on the next screen after generating the HuBMAP IDs


Description

Metadata [+ Add Metadata](#)

Image [+ Add Image](#)

Make sure any uploaded images are de-identified

[Generate ID](#) [Cancel](#)



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HuBMAP Display ID Generator

Generate unique identifiers which will be used consortium wide to track sample and associate data with samples.

3 sample ids were generated: TEST0005-RK-6 through TEST0005-RK-8

Type: FFPE block

[Assign Lab IDs and Sample Locations](#)

[Return to Search](#)


Assign Lab IDs and Sample Location

Lab Sample Id	Register Location	SuccessView JSON
TEST0005-RK-6	<input type="text" value="TEST0005-RK-6-A"/> Register Location	
TEST0005-RK-7	<input type="text"/> Register Location	
TEST0005-RK-8	<input type="text"/> Register Location	

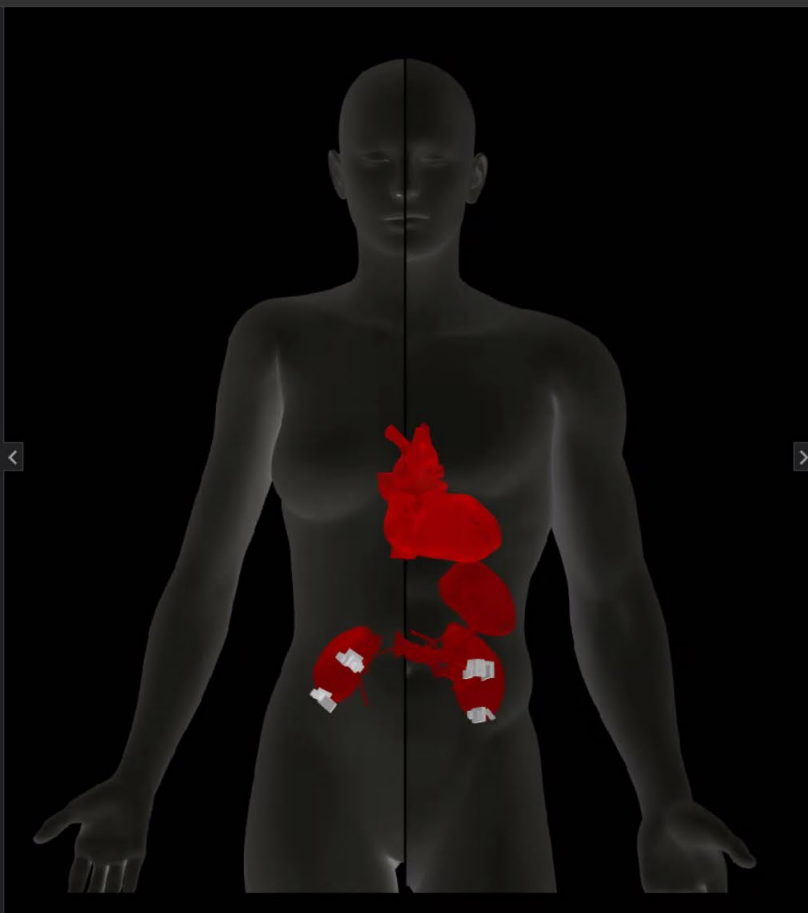
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Thanks go to the IEC for providing screenshots

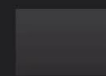
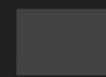
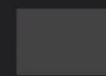
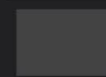
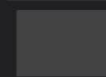
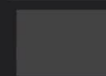
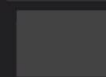
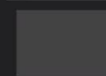
Search ontology terms ... 

- body
 - heart
 - lung
 - kidney
 - right kidney
 - left kidney
 - kidney capsule
 - cortex of kidney
 - renal medulla
 - renal column
 - renal pyramid
 - hilum of kidney
 - kidney interstitium
 - kidney calyx
 - renal pelvis
 - ureter
 - renal papilla
 - renal fat pad
 - nephron
 - spleen
 - colon
 - small intestine
 - rectum



Kidney

0 Donors
1867 Samples
4 Datasets

- 
VAN0023-LK-1: Female, Age 44
 TMC-Vanderbilt
 2/5/28
 Sample
- 
VAN0012-RK-103-93: Female, Age 44
 TMC-Vanderbilt
 10/24/19 - 2D sections: 75-80, Imaging odd, MxIF e...
 Sample
- 
VAN0012-RK-103-83: Female, Age 44
 TMC-Vanderbilt
 10/24/19 - 2D sections: 75-80, Imaging odd, MxIF e...
 Sample
- 
VAN0012-RK-103-95: Female, Age 44
 TMC-Vanderbilt
 10/24/19 - 2D sections: 75-80, Imaging odd, MxIF e...
 Sample
- 
VAN0012-RK-103-52: Female, Age 44
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 10/24/19 - 2D sections: 75-80, Imaging odd, MxIF e...
 Sample
- 
VAN0012-RK-103-3: Female, Age 44
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 10/24/19 - 2D sections: 75-80, Imaging odd, MxIF e...
 Sample
- 
VAN0012-RK-103-19: Female, Age 44
 TMC-Vanderbilt
 10/24/19 - 2D sections: 75-80, Imaging odd, MxIF e...
 Sample
- 
VAN0012-RK-103-68: Female, Age 44
 TMC-Vanderbilt
 10/24/19 - 2D sections: 75-80, Imaging odd, MxIF e...
 Sample

1st Portal Release

Search | HuBMAP

localhost:5000/search?entity_type[0]=Dataset

Donors Samples Datasets **CCF-UI** Help Logout

Search

Search

Creator

- Joel Welling 116
- Desheng Li 56
- Chuck Borromeo 30
- Jesus Penaloza Aponte 23
- Nico Pierson 18

[View all](#)

Contains genetic sequences

- no 211
- yes 30

Data type

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- ["dataset", "codex", "cytokit"] 13
- ["scRNA-Seq-10x"] 11

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Type: Dataset x Clear all filters

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	HBM356.QRPV.263	Lymph Node IMC	
	HBM976.LGXF.476	Test Dataset 24	
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	HBM368.MQGB.483		
	HBM257.ZNBS.437		
	HBM533.PRKM.985		
	HBM396.RPRR.624	Case 4 Spleen 2	
	HBM667.HNSN.549		
	HBM972.GWWH.687		

Early draft - limited functionality and design until more data has been indexed.

ASCT Table Design

The CCF Session at the NIH-HCA meeting—co-organized with Peter Hunter (SPARC) and James Gee (BICCN)—brought together experts across consortia.

In follow up meetings, 10 ASCT tables have been created via collaborations across consortia. Ontology experts, including Chris Mungall and Mark Musen, provided expert comments.



	HuBMAP	RBK	KPMP	SPARC	LungMAP	HTAN	HCA	GUDMAP	Gut Cell Atlas	BICCN	Allen Brain	TCGA	Wellcome	MRC	H2020	GTEx	Total
Kidney	1	1	1	0	0	0	1	1	0	0	0	1	1	1	0	1	9
Liver	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	1	3
Spleen	1	0	0	0	0	0	1	0	0	0	0	0	1	0	0	1	4
Heart	1	0	0	1	0	0	1	0	0	0	0	0	0	0	0	1	4
Lung	1	0	0	1	1	1	1	0	0	0	0	1	1	1	1	1	10
L intestine/Colon	1	0	0	1	0	1	1	0	1	0	0	1	0	0	0	1	7
S intestine	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	2
Bladder	1	0	0	1	0	0	0	1	0	0	0	1	0	0	0	1	5
Ureters	1	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	2
Thymus	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	2
Lymph nodes	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	2
mediastinal lymph node	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1
Eye	1	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	3
Brain	0	0	0	0	0	0	1	0	0	1	1	1	0	0	1	1	6
Brain stem	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1
Cerebellum	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	1	3
Spinal cord	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	2
Pancreas	0	0	0	0	0	1	1	0	0	0	0	1	0	0	1	1	5
Breast	0	0	0	0	0	1	1	0	0	0	0	1	1	0	0	1	5
Skin	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	1	3
Pediatric systems	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0	2
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Cervix	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1
Uterus	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	5
Blood	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0	2
Bone	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1
Placenta	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1
Decidua	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1
Embryo	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1
esophagus	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	1	3
hematopoietic system	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	2
immune system bulk	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1
Stomach	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	1	3
Thyroid	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	2
Prostate	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	1	3
Adrenal gland	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	3
Totals	11	1	1	7	1	6	21	4	1	2	2	20	7	5	4	21	114

<https://tinyurl.com/ASCT10x10>

<https://www.humancellatlas.org/coordinators>



ASCT Table Meetings

Meetings take place monthly to

- Review and approve tables.
- Formalize and unify table design language.
- Discuss table usage.

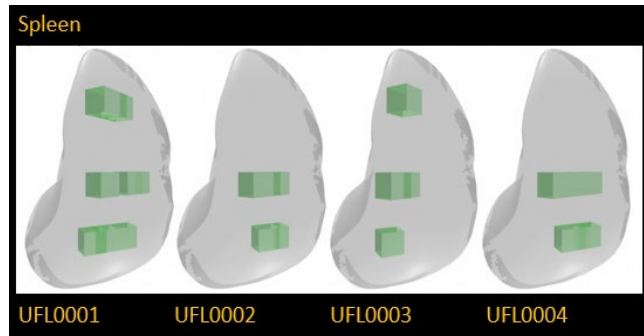
We are working on

- Converting tables into machine readable formats, see example for vasculature.
- Compare tables against Uberon, CL, and other ontologies.
- Compare tables against cell types identified in harmonized HuBMAP data and data generated by other efforts.

ASCT Table Usage

Compare cell types in ASCT tables with cell types identified in HuBMAP data.

Spleen example: Data from five tissue blocks from 4 spleens were harmonized.



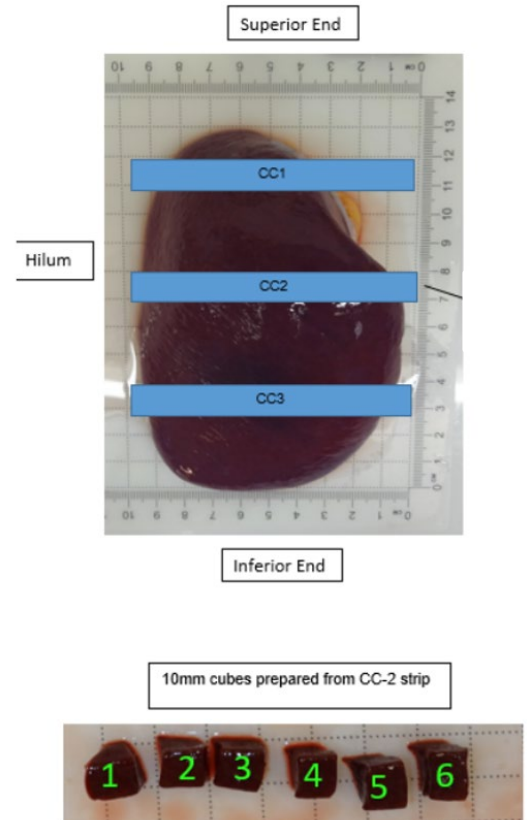
14 years
Female

11m
Male

18y
Male

18y
Male

- UFL0001-SP-2-8, cube 1
- UFL0001-SP-3-4, cube 3
- UFL0002-SP-2-2, cube 3
- UFL0003-SP-2-2, cube 1
- UFL0004-SP-2-1, cube 4



ASCT Table Usage

Seurat harmonization results: Cell counts and prediction scores

UFL0001-SP-3-4, cube 3	UFL0003-SP-2-2, cube 1	UFL0002-SP-2-2, cube 3	UFL0001-SP-2-8, cube 1	UFL0004-SP-2-1, cube 4
HBM336.FWTN.636	6010 HBM396.RPRR.624	9382 HBM472.NTNN.543	8738 HBM556.QMSM.776	5273 HBM984.GRBB.858
alpha-beta T cell	372	773	1497	515
B cell	1349	4463	6550	1407
CD141-positive myeloid dendritic cell	55	89	19	44
CD14-positive monocyte	1851	2242	238	872
CD1c-positive myeloid dendritic cell	185	18		162
erythroblast	177	42	17	89
gamma-delta T cell	151	57	30	176
hematopoietic stem cell	73	84	56	79
low-quality	93	71	94	129
natural killer cell	594	1307	84	460
plasma cell	460	171	101	360
plasmablast	22	7	28	1
splenic endothelial cell	424	47	6	588
splenic fibroblast	15	5	6	20
splenic macrophage	189	6	12	371
				6328
				878
				2803
				17
				802
				19
				1241
				30
				131
				267
				75
				36
				1
				2
				26

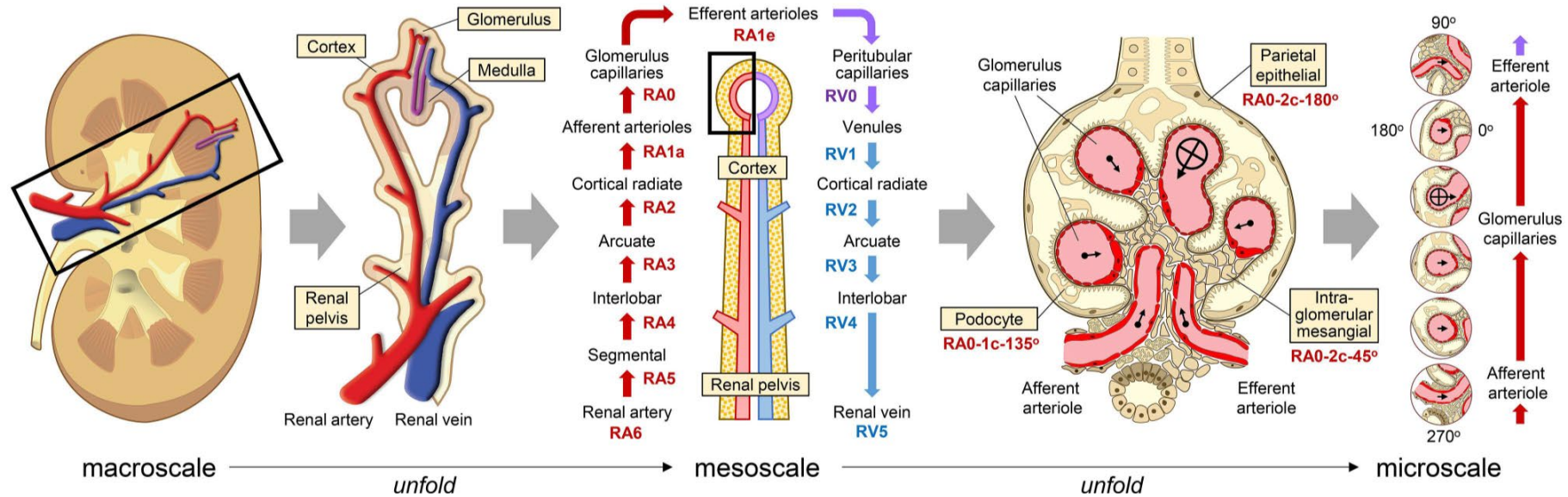
HBM336.FWTN.636	1 HBM396.RPRR.624	HBM472.NTNN.543	HBM556.QMSM.776	HBM984.GRBB.858
alpha-beta T cell	1	0.81531392	0.935717083	0.852839869
B cell	1	0.966724409	0.978509075	0.958394632
CD141-positive myeloid dendritic cell	1	0.714928889	0.870885148	0.890455775
CD14-positive monocyte	1	0.940118127	0.937989865	0.938657155
CD1c-positive myeloid dendritic cell	1	0.563719677		0.803740621
erythroblast	1	0.532657229	0.694290983	0.978894926
gamma-delta T cell	1	0.539497766	0.502216796	0.740586674
hematopoietic stem cell	1	0.726385726	0.764763311	0.75724491
low-quality	1	0.680657174	0.68445931	0.649134708
natural killer cell	1	0.770988817	0.623959124	0.833128238
plasma cell	1	0.945743141	0.947816498	0.954210246
plasmablast	1	0.445989963	0.593308367	0.535441087
splenic endothelial cell	1	0.921750546	0.649250648	0.949515886
splenic fibroblast	1	0.883399167	0.886345281	0.878256094
splenic macrophage	1	0.646221909	0.827945537	0.853436501
				0.819379533
				0.963947327
				0.615445273
				0.963592885
				0.783471674
				0.904053159
				0.648865012
				0.728998052
				0.718629642
				0.82474102
				0.558989332
				0.555504115
				0.820886903
				0.880481635

Example: Converting tables into machine readable formats – Kidney vasculature

Vasculature	renal artery [L/R]	segmental arteries [superior, inferior, anterior, posterior]		Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>		
		interlobar arteries						
		arcuate arteries						
		cortical radiate arteries {cortex}						
		{nephron}	afferent arterioles		EC-Afferent/Efferent Arteriole	EC-AEA	<i>SERPINE2*</i> , <i>TM4SF1*</i>	
			glomerulus capillaries {glomerulus}	Capillary Endothelial Cell		GC-EC	<i>EHD3*</i> , <i>EMCN*</i> , <i>HECW2*</i> , <i>FLT1*</i> , <i>AQP1*</i>	
				efferent arterioles		EC-Afferent/Efferent Arteriole	EC-AEA	<i>SERPINE2*</i> , <i>TM4SF1*</i>
				peritubular capillaries	EC-Peritubular capillaries		EC-PTC	<i>PLVAP*</i>
					descending vasa recta		EC-Descending Vasa Recta	EC-DVR
		ascending vasa recta			EC-Ascending Vasa Recta	EC-AVR	<i>DNASEIL3*</i>	
renal vein [L/R]	cortical radiate veins {cortex}		Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>			
	arcuate veins							
	interlobar veins							
	venules {nephron}							

Vasculature	renal artery [L/R]			Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	
Vasculature	renal artery [L/R]	segmental arteries [superior, inferior, anterior, posterior]		Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	
Vasculature	renal artery [L/R]	interlobar arteries		Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	
Vasculature	renal artery [L/R]	arcuate arteries		Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	
Vasculature	renal artery [L/R]	cortical radiate arteries {cortex}		Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	
Vasculature	renal artery [L/R]	cortical radiate arteries {cortex}	afferent arterioles {nephron}	EC-Afferent/Efferent Arteriole	EC-AEA	<i>SERPINE2*</i> , <i>TM4SF1*</i>	
Vasculature	renal artery [L/R]	cortical radiate arteries {cortex}	afferent arterioles {nephron}	glomerulus capillaries {glomerulus}	Capillary Endothelial Cell	GC-EC	<i>EHD3*</i> , <i>EMCN*</i> , <i>HECW2*</i> , <i>FLT1*</i> , <i>AQP1*</i>
Vasculature	renal artery [L/R]	cortical radiate arteries {cortex}	efferent arterioles {nephron}	EC-Afferent/Efferent Arteriole	EC-AEA	<i>SERPINE2*</i> , <i>TM4SF1*</i>	
Vasculature	renal artery [L/R]	cortical radiate arteries {cortex}	efferent arterioles {nephron}	peritubular capillaries	EC-Peritubular capillaries	EC-PTC	<i>PLVAP*</i>
Vasculature	renal artery [L/R]	cortical radiate arteries {cortex}	efferent arterioles {nephron}	descending vasa recta	EC-Descending Vasa Recta	EC-DVR	<i>TM4SF1*</i> , <i>PALMD</i>
Vasculature	renal artery [L/R]	cortical radiate arteries {cortex}	efferent arterioles {nephron}	ascending vasa recta	EC-Ascending Vasa Recta	EC-AVR	<i>DNASEIL3*</i>
Vasculature	renal vein [L/R]	cortical radiate veins {cortex}	venules {nephron}	Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	
Vasculature	renal vein [L/R]	cortical radiate veins {cortex}		Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	
Vasculature	renal vein [L/R]	arcuate veins		Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	
Vasculature	renal vein [L/R]	interlobar veins		Endothelial Cell (general)	EC	<i>EMCN*</i> , <i>PECAM1*</i> , <i>FLT1*</i>	

Capturing vasculature details is critically important for a vasculature based CCF



Weber, Griffin M, Yingnan Ju, and Katy Börner. 2020. "[Considerations for Using the Vasculature as a Coordinate System to Map All the Cells in the Human Body](https://doi.org/10.3389/fcvm.2020.00029)". *Frontiers in Cardiovascular Medicine* 7 (29): doi: 10.3389/fcvm.2020.00029.



ASCT Table Discussion

Please join the CCF breakout for a discussion of

- Existing tables
- Planned analyses and visualizations of ASCT+B tables
- Table usage for research design and quality control

Contact MC-IU via infoccf@indiana.edu for questions and suggestions.

Apply for a **Postdoc position** to advance ASCT+B curation and usage, e.g., to understand vascular pathways in the human body (arteries, veins, capillaries, and lymph vessels).

Acknowledgements

HuBMAP Consortium (<https://hubmapconsortium.org>)



Thanks go to all the **patients** that agreed to volunteer healthy tissue and open use of their data.



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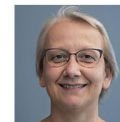
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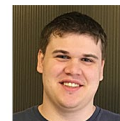
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